

This endeavor sought to determine the hormonal mechanism driving post-ecdysial shell calcification in crustaceans. Carbonic anhydrase (CA) catalyzes the carbon dioxide hydration reaction, which generates bicarbonate ions essential for calcium carbonate formation. Using metal analysis, enzymatic assay, tissue culture, and real-time PCR, my lab has found that epidermal CA activity is correlated to exoskeletal calcium content and that both epidermal CA activity and CA gene expression are inducible by the molting hormone in the blue crab *Callinectes sapidus*. These results have led to the new paradigm of postecdysial shell mineralization being controlled by the molting hormone. Intriguingly, while molting hormone injection elevated epidermal CA activity, the exogenous molting hormone was found to suppress shell calcification, implicating that sclerotization and mineralization of the new shell must be coordinated and enhanced deposition of calcium carbonate as a result of increased epidermal CA activity following hormonal treatment would be inhibited to avoid the formation of a structurally defective exoskeleton.

Background

• Crab shell, a biological concrete, is made up of an organic matrix of chitin and proteins ("steel rods") mineralized by inorganics, primarily carbonate salts ("cement").

• After molting, crustaceans are covered by a soft shell, which must harden rapidly. Shell hardening consists of sclerotization ("tightening of steel rods") and mineralization ("addition of cement").



Г Chitin (glucosamine polymer)– Proteins (such as collagens) Inorganics (such as CaCO₃) -



Scientific Question to Be Addressed

While it well known that sclerotization is controlled by the neurohormone bursicon [1], the hormonal mechanism driving mineralization remains unresolved. This project attempted to address the question of which hormone controls mineralization in crustaceans, using the blue crab, Callinectes sapidus, as the model crustacean.

Shell hardening

Sclerotization or Tanning of organic matrix ("Tightening of steel rods")

Controlled by ???? ("Cement addition")

Hypothesis

Given the similarity between the patterns of changes in molting hormone concentration and epidermal carbonic anhydrase (CA) activity during the molting cycle [2], it was hypothesized that the molting hormone regulates mineralization through controlling epidermal CA due to the following reactions.

Carbonic Anhydrase

 $CO_2 + H_2O$

 $HCO_{3}^{-} + Ca^{2+}$

Hormonal Control of the Making of Crab Shell, a Biological concrete

Enmin Zou Department of Biological Sciences, Nicholls State University

Abstract

Controlled by bursicon

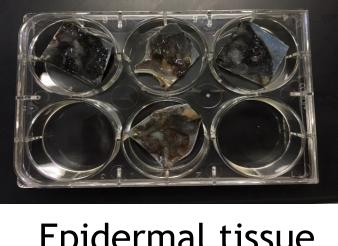
 $H^+ + HCO_3^-$

+ $CaCO_3$

laboratory.



A molting crab



2. Calcium content in the shell was analyzed using ICP-OES. 3. Epidermal CA activity was assayed according to Calhoun and Zou [2].

4. The in vitro molting hormone 20-hydroxyecdsone (20-HE) treatment of epidermal tissues followed Booth and Zou [3], while for in vivo treatment 20-HE was injected to postecdysial crabs.

5. The expression of epidermal CA genes (CA mRNA) was quantified using quantitative real-time PCR. 6. Epidermal CA gene expression of knocked down using RNAi (through injection of dsRNA to softshell crabs).

Results and Discussion

1. Significant correlation between epidermal CA activity and calcium content (Fig. 1) suggests that CA mediates calcium incorporation into the exoskeleton.

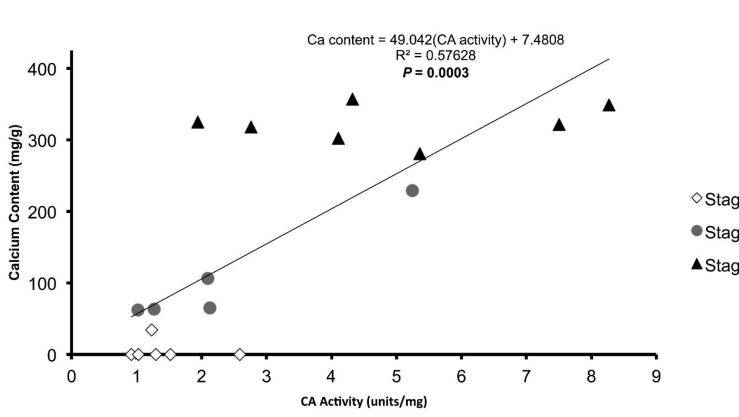


Figure 1. Correlation between epidermal CA activity and exoskeletal calcium content in post-ecdysial blue crabs.

2. Significant in vitro induction of epidermal CA mRNA (Fig. 2) and significant in vivo induction of epidermal CA activity (Fig. 3) suggest that epidermal CA is controlled by and hemocyte granulation in the blue crab, Callinectes sapidus. PLoS One 7(9), e46299. the molting hormone.

Methodologies

1. The "buster" blue crabs purchased from local softshell crab makers were used to produce postmolt crabs in the



Epidermal tissue culture

ICP-OES for metal analysis

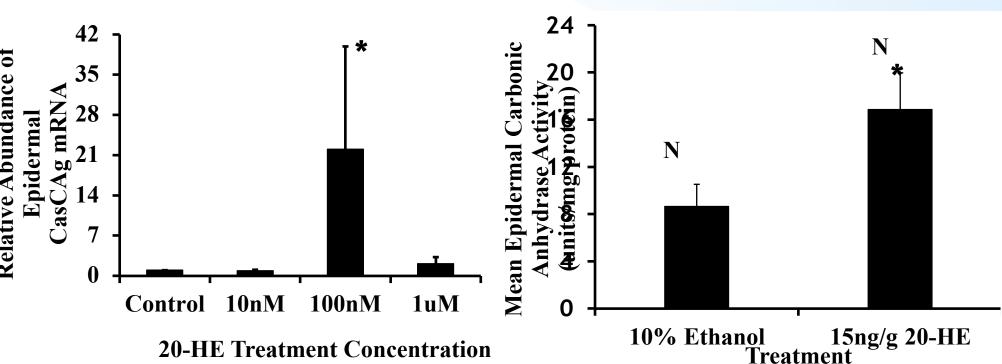


Figure 2. In vitro induction of epidermal CA gene (CasCAg mRNA) by 20-HE. N = 5, * P < 0.05 relative to control.

3. Interestingly, injection of the exogenous molting hormone inhibited shell calcium content (Fig. 4).

This result implicates that sclerotization and mineralization of the new shell must be coordinated and enhanced deposition of calcium carbonate as a result of increased epidermal CA activity following hormonal treatment would be inhibited to avoid the formation of a structurally defective exoskeleton.

4. Preliminary RNAi results

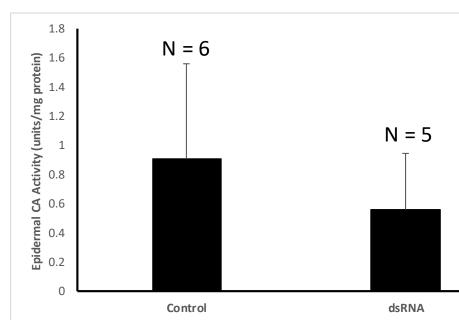


Figure 5. Knock-down of epidermal CA activity by 10 µg *CasCAg* dsRNA/crab.

Epidermal CA is controlled by the molting hormone, suggesting that the molting hormone is involved in the regulation of postecdysial shell mineralization in Crustacea.

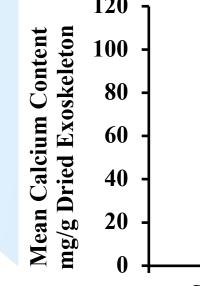
Future Investigation

Further RNAi experiments will be conducted to ascertain the role of epidermal CA in post-ecdysial deposition of calcium to the shell.

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[1] Chung, J.S., Katayama, H., Dircksen, H., 2012. New functions of arthropod bursicon: inducing deposition and thickening of new cuticle [2] Calhoun, S., Zou, E. (2016) Epidermal carbonic anhydrase activity and exoskeletal metal content during the molting cycle of the blue crab, Callinectes sapidus. J. Exp. Zool. 325A:200-208. [3] Booth, A., Zou, E. (2016) Impact of molt-disrupting BDE-47 on epidermal ecdysteroid signaling in the blue crab, *Callinectes sapidus*, in vitro. Aquatic Toxicology 177:373-379.





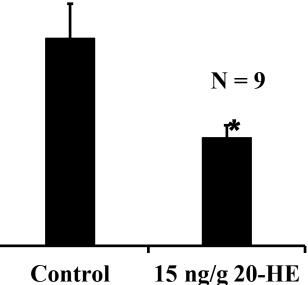
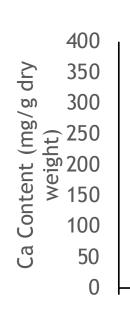


Figure 3. In vivo induction of epidermal CA activity

by 20-HE. * P < 0.05.

Figure 4. Effect of 20-HE injection on shell calcium content. * P < 0.05.



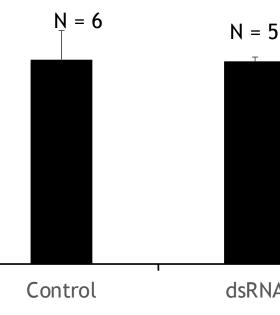


Figure 6. Effect of CasCAg dsRNA injection on exoskeletal Ca content.

Conclusion

Acknowledgements